

Howto T-REX FlowJo Plugin

Introduction

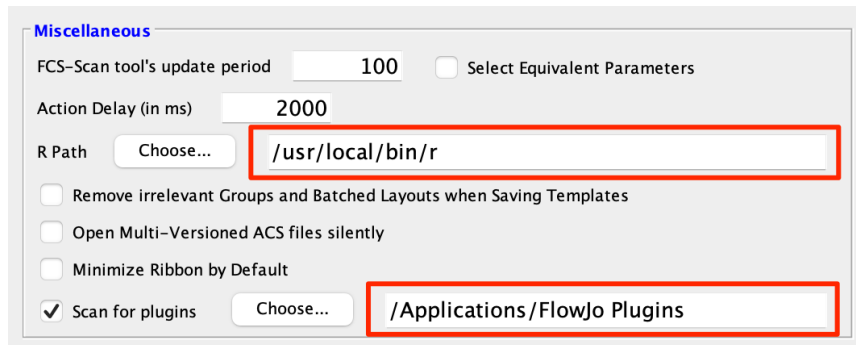
The Tracking Responders Expanding (T-REX) algorithm has been implemented as a FlowJo plugin. The T-REX algorithm creates a pair of dimensionality reduction parameters, identifies the areas of difference between two user specified conditions, clusters the data, then identifies the clusters that match the region of difference.

The T-REX FlowJo plugin is designed to be run on a single FCS file that contains two gates identifying events that were processed under different experimental conditions. Such an FCS file can be created by concatenating two or more individual FCS files, and adding a keyword-based parameter that defines which samples belong to a given experimental condition. Please refer to this documentation page on concatenation for more details: <https://docs.flowjo.com/flowjo/workspaces-and-samples/samples-and-file-types/ws-export/>

The plugin will create 4 new derived parameters added to the selected file in the FlowJo workspace - two dimensionality reduction parameters, one cluster results parameter, and one parameter that measures difference between conditions, along with gated cluster populations if differences are identified. Additional results files produced by the algorithm will be found within a TRex output_files folder within the derived parameters folder for the workspace in which the plugin was run.

Download and Installation

1. Place the plugin.jar file in your Plugins folder, and direct FlowJo to that folder using the Diagnostics section of the Preferences.
2. Make sure you have R installed and the R path is specified in the R Path field of the Diagnostics section of the Preferences.



3. Restart the FlowJo application to pick up the new plugin.

Note: Running the T-REX plugin for the first time will install the R package dependencies required to run the T-REX algorithm into your R library. To install manually, type (or copy/pasting) the following in your R console:

```
install.packages(c('tidyverse','ggplot2','cowplot','FNN','dbscan','fancycut','gridExtra','uwot',  
                  'RColorBrewer','dplyr','purrr','Rtsne','BiocManager'))  
BiocManager::install(c('cytoMEM','flowCore','Biobase'))
```

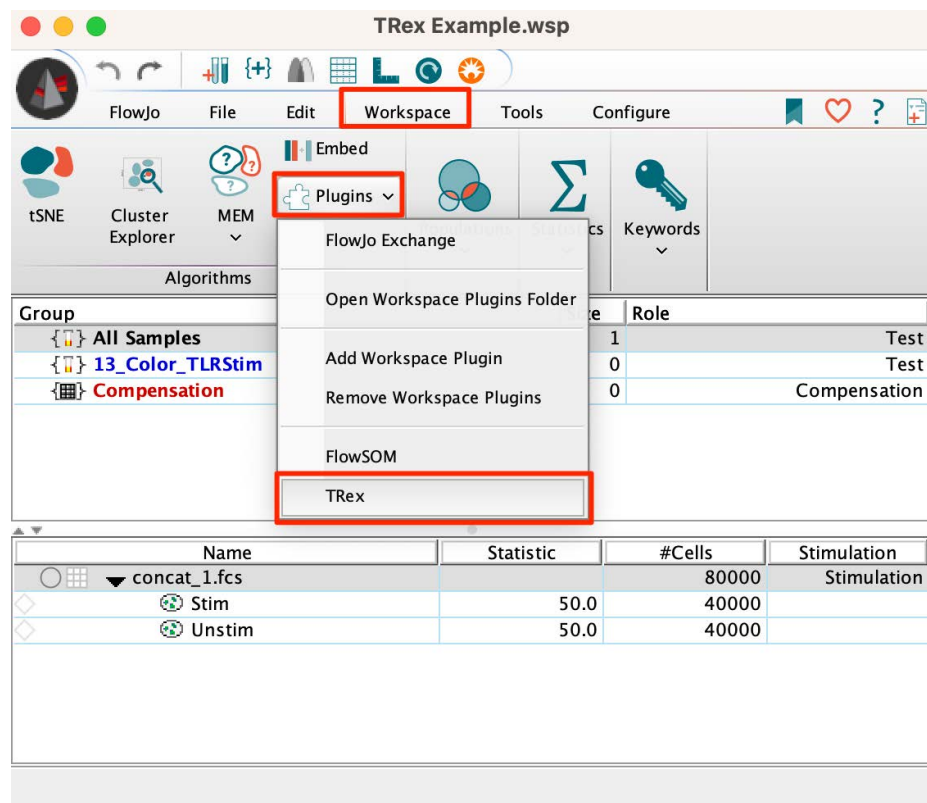
For older versions of R, please refer to the appropriate Bioconductor release: <https://bioconductor.org/about/release-announcements/>

Additional documentation on installing plugins in FlowJo can be found here: <http://docs.flowjo.com/d2/plugins/installing-plugins>.

Usage

To run the T-rex plugin on your FCS file:

1. Select a sample of interest with 2 gates identifying 2 populations to compare within the workspace.
2. Go to the Workspace tab and select the TRex option from within the Plugins dropdown menu.



3. This dialog will appear:

TRex (1.1.0)

Comp-APC-A :: IL-6
Comp-APC-H7-A :: HLA-DR
Comp-Alexa Fluor 430-A :: Via
Comp-Alexa Fluor 700-A :: CD1c
Comp-BUV396-A :: TNF
Comp-BUV737-A :: CD83
Comp-FITC-A :: CD14 Ax488
Comp-PE-A :: IL-1B
Comp-PE-Cy7-A :: CD3-56-19-20
Comp-PE-Texas Red-A :: CD16 PE-CF594
Comp-Pacific Blue-A :: IL-12 BV421
Comp-PerCP-Cy5.5-A :: CD123

Pre-Treatment population: Unstim

Post-Treatment population: Stim

K: 60

eps: 0.30

Bins: [0,5],[5,15],[15,85],[85,95],[95,100]

Comparison method: UMAP

t-SNE Perplexity: 30

t-SNE Iterations: 1000

Minimum Cluster Size: 10

If you are using this plugin, please cite:
<https://elifesciences.org/articles/64653>

Cancel OK

In the top box you must select the parameters for the algorithm to consider. In general, select any parameter that may have differentiating power between the groups. In this example, the live/dead and dump channels have been omitted as those parameters were employed for gating during pre-processing. The top two selection boxes must be used to select the two populations to compare, using gates as the method for identifying cells of each type.

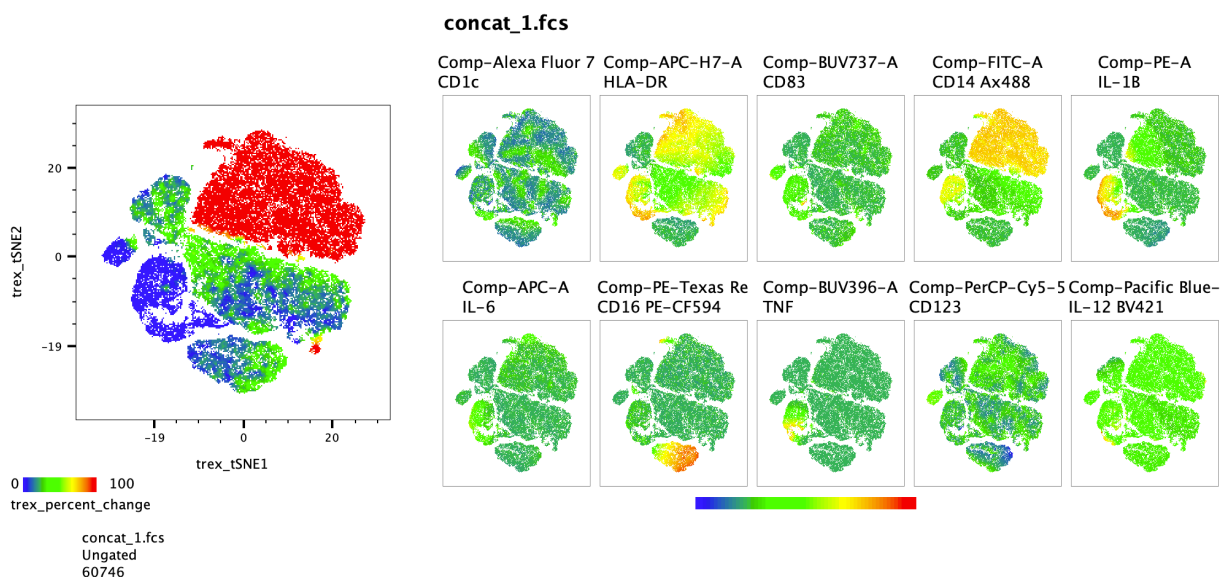
Other options include:

- **K** - The number of nearest neighbors to consider during clustering.
- **eps** - The radius of the epsilon neighborhood. Two points are considered neighbors if the distance between them is below the threshold epsilon.
- **Bins** - The percentage of cells from one condition to be highlighted.
- **Comparison method** - UMAP or tSNE
- **t-SNE perplexity** - Size of the neighborhood setting, exclusively for the tSNE choice.
- **t-SNE iterations** - Set a maximum limit on tSNE iterations.
- **Minimum Cluster Size** - Minimum number of events to be considered a cluster

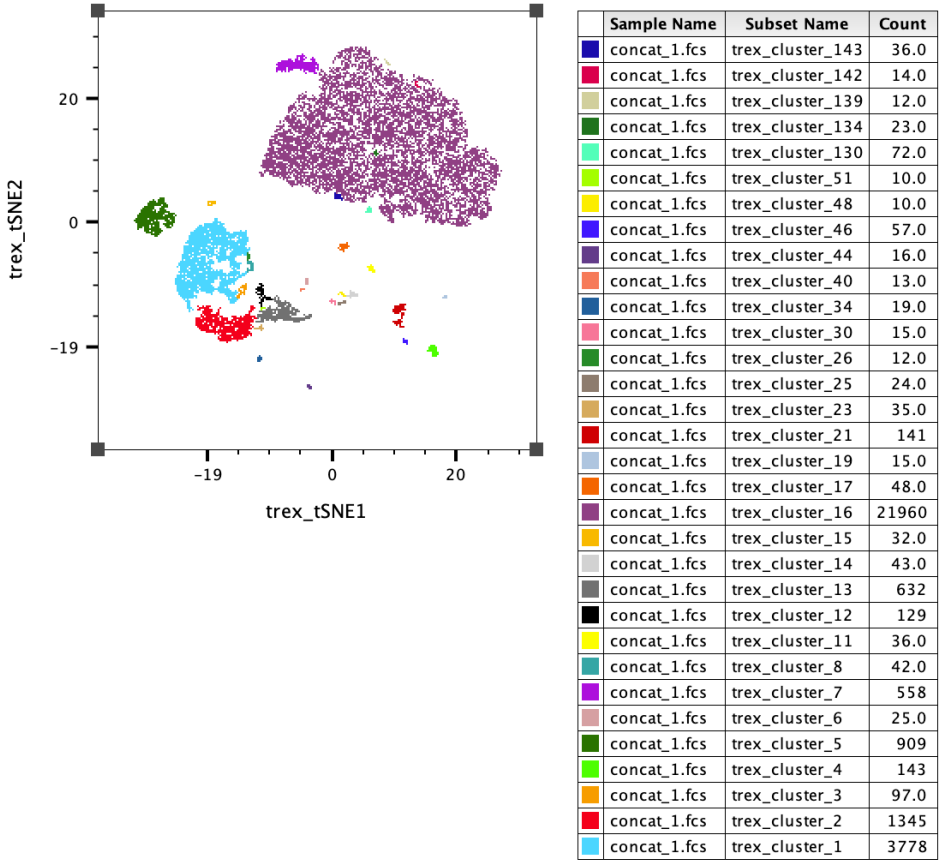
The results will be four derived parameters: two `trex_DRmethod` parameters representing the low dimensional embedding results using the selected dimensionality reduction method, a `trex_percent_change` parameter (a measure of the representation of each class across the dimensionality reduction space), and a `trex_cluster` parameter for identifying clusters.

Name	Statistic	#Cells
▼ concat_1.fcs		60746
% trex_cluster		
% trex_percent_change		
% trex_tSNE1		
% trex_tSNE2		
TREx		
Stim	50.0	30373
trex_cluster_1	6.22	3778
trex_cluster_2	2.21	1345
trex_cluster_3	0.16	97
trex_cluster_4	0.24	143
trex_cluster_5	1.50	909
trex_cluster_6	0.041	25
trex_cluster_7	0.92	558
trex_cluster_8	0.069	42

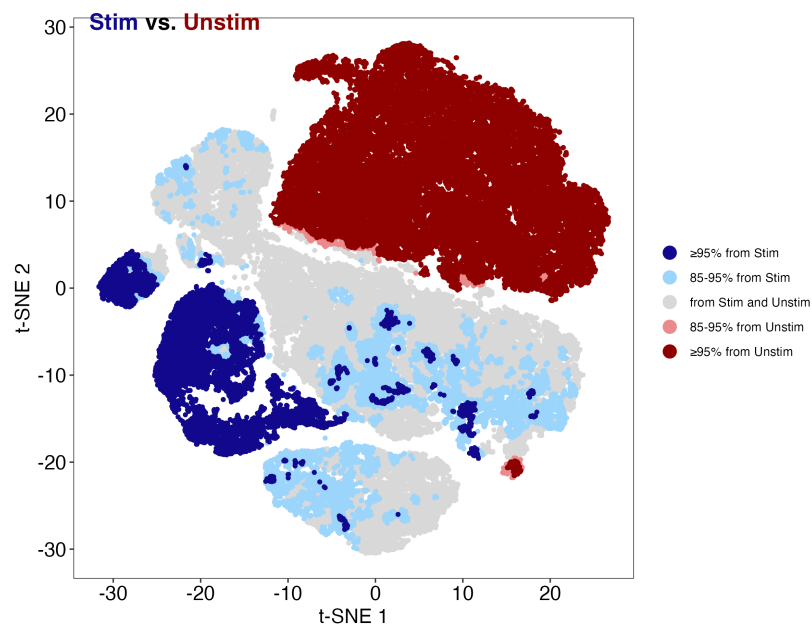
Plotting this outcome using the tSNE parameters and heat mapping by `tex_percent_change`, then juxtaposing it with a heat map of parameter expression can provide an understanding of the data and where it varies between conditions.



In this example we can see that the unstimulated cells for a population that is largely unique from the stimulated cells, appearing bright red in the plot. And likewise, the stimulated cells for a couple unique blue populations. The cluster results can also be viewed in FlowJo, perhaps most usefully as an overlay. The plot below shows the clusters with differential power.



A number of useful plots are created in the TRex derivative folder as well. Perhaps the most informative is the blue-red cluster plot that shows which regions have significant expansion of one category or the other:



Leave us your feedback

Please write to flowjo@bd.com with any questions or concerns.

References

1. Sierra M Barone, Alberta GA Paul, Lyndsey M Muehling, Joanne A Lannigan, William W Kwok, Ronald B Turner, Judith A Woodfolk, Jonathan M Irish. *Unsupervised machine learning reveals key immune cell subsets in COVID-19, rhinovirus infection, and cancer therapy.*